Causal pathogen: Ascochyta rabiei

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- The pathogen is introduced through infected seed and via airborne inoculum. Note that infected seeds are often symptomless.
- Timing of disease onset depends on inoculum arrival, but disease symptoms often appear at the late vegetative stages or bloom initiation.
- Infections result in tan to brown lesions on leaves, stems, and pods.
- Within lesions, concentric rings of small gray to brown specks can generally be found. Each of these specks is a pycnidium, a tiny flask-shaped fruiting structure containing thousands of spores of the causal pathogen.
- Diseased pods often fail to produce seed or may produce shriveled, discolored seeds.
- · Ascochyta blight can reach epidemic levels very quickly.
- Even low levels of foliar disease during the bloom period can lead to high levels of pod infections during the pod-fill period. Because many infected pods do not set seed, management of pod infections is critical for preserving yield potential and maintaining seed quality.
- Disease risk is always highest during periods of rainfall and/or heavy dews.

FOLIAR AND POD LESIONS:



STEM LESIONS:













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1. Select a moderately resistant variety.

- B-90 (also known as Amit) is a small-seeded Kabuli chickpea with moderate resistance to Ascochyta blight.
- CDC Frontier is a medium-seeded Kabuli chickpea with moderate resistance to Ascochyta blight.
- CDC Luna is a large-seeded Kabuli chickpea with fair resistance to Ascochyta blight.

2. Use seed that has been tested for Ascochyta.

- · Ascochyta is seed-borne, and the disease is readily transmitted from infested seeds to seedlings.
- Diseased seeds are not always discolored. Make sure your seed has been tested for Ascochyta; the plant disease diagnostic labs at NDSU and MSU conduct this test. It is best to only use seed that tests completely negative for Ascochyta. Never use seed with Ascochyta incidence greater than 0.3%.

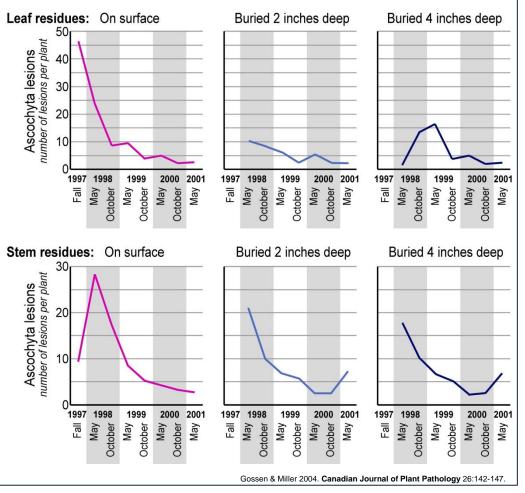
3. Treat your seed with the fungicide thiabendazole (Mertect 340-F).

- Treat your seed with thiabendazole even if your seed test indicates an incidence of Ascochyta infection of 0%. Seed testing is generally done on a relatively small seed sample (500 seeds), and few seed lots are completely free of Ascochyta-infected seeds.
- Other commonly used seed treatments (eg Apron Maxx) are not very effective against Ascochyta. Likewise, thiabendazole is not very
 effective against Phytophthora root rot and other seed and seedling diseases of chickpea. Treat with a mix of thiabendazole and other
 products if you need protection against seed and seedling diseases.

4. <u>Long rotations out of</u> <u>chickpeas are best.</u>

- A minimum 3-year rotation out of chickpeas is recommended.
- Low levels of Ascochyta transmission from chickpea residues to a new crop may occur even after rotating out of chickpeas for 3 years. Research conducted in a heavy clay soil in Saskatoon, SK suggests that transmission from infested chickpea residues to new chickpeas drops sharply after a 2-year rotation out of chickpeas but can continue for at least 4 years since the previous crop. The effect of soil type and climate on survival of the Ascochyta pathogen is poorly understood, and it is unknown whether different results might be obtained in different soil types or climates.
- In addition to small grains and oilseed crops, lentils and field peas are acceptable rotational crops. Ascochyta rabiei, cause of Ascochyta blight on chickpeas, is specific to chickpeas; other Ascochyta species cause Ascochyta blight on lentils and field peas.

Transmission of Ascochyta from infested chickpea leaf and stem residues to new chickpea plants may occur for at least 4 years. Numbers represent the average number of Ascochyta lesions that developed in greenhouse-grown chickpea seedlings inoculated with chickpea residues recovered from a site near Saskatoon, SK. Residues were buried 0, 2, or 4 in. deep in a heavy clay soil.



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5. Use foliar fungicides.

- This disease can reach epidemic levels very quickly, and fungicide applications must be made in a timely manner.
- When possible, a preventative approach to managing this disease is recommended:
 - If no disease has been detected 7 to 10 days before bloom initiation (if there have been frequent rain events and/or heavy dew) or at bloom initiation (if it is dry), a preventative application of chlorothalonil (Bravo WS, Echo 720, etc) is advised.
 - Seven to ten days after applying chlorothalonil, an application of a systemic fungicide (Proline, Endura, Priaxor, etc.) is generally advised.
 - If the weather remains favorable for disease, sequential applications of fungicides should continue every 10 to 14 days during the critical bloom and pod-fill period.
- Ascochyta blight often develops at the mid- to late vegetative stages before bloom.
 - Chickpea crops are at highest risk of early Ascochyta blight development when (1) they are located downwind of fields where chickpea Ascochyta blight occurred the previous season or (2) seed with seed-borne Ascochyta is planted. The Ascochyta pathogen produces airborne spores on overwintered diseased crop residues, and these spores are capable of long-distance movement in the atmosphere.
 - Ascochyta blight often first appears as scattered disease lesions on individual plants.
 - If the weather is cool and wet, significant spread from these foci of infection can occur before bloom; otherwise, the disease may not spread until the canopy closes.
 - In order to prevent secondary spread of Ascochyta blight from the initial foci of infection, applications of systemic fungicides are
 often advised as soon as trace levels of disease are detected, particularly if the weather is favorable for disease and/or the crop is
 nearing canopy closure. If the weather remains favorable for disease, sequential applications of fungicides should continue every
 10 to 14 days during the critical bloom and pod-fill period.
- Even if disease is at low levels at the end of bloom, a fungicide application may be warranted during the pod-fill stage. Even when foliar disease is at low levels, the incidence of pod infection can be quite high. When pods become infected, they often do not produce seeds.
- 6. Selecting fungicides for Ascochyta management on chickpeas
- Rotation fungicide chemistries is critical. Inadequate rotation of fungicide chemistries caused a loss of efficacy of Qol (strobilurin/FRAC 11) fungicides (Headline and Quadris).
- Rotate between DMI (FRAC 3) and SDHI (carboxamide; FRAC 7) fungicides
 - <u>DMI FUNGICIDES</u>: **Proline** (prothioconazole) is registered (5.0 and 5.7 fl oz/ac)
 - <u>SDHI (CARBOXAMIDE) FUNGICIDES</u>:
 <u>Endura</u> (boscalid) is registered (6 oz/ac)
 <u>Priaxor</u> (fluxapyroxad + pyraclostrobin) is registered (4 to 6 fl oz/ac).
 Because pyraclostrobin is no longer effective against Ascochyta blight of chickpea, you can assume that disease control conferred by Priaxor is primarily due to the carboxamide ingredient, fluxapyroxad.
 Vertisan (penthiopyrad) is registered (14 to 20 fl oz/ac).
- Consider tank-mixing DMI or SDHI (carboxamide) fungicides with chlorothalonil (Bravo WS, Echo 720, etc.), especially if the chickpea canopy is still open. Chlorothalonil has a different mode of action (method of controlling fungi) than other fungicides, and the use of chlorothalonil will help delay the development of fungicide resistance.
- Strobilurin (Qol/FRAC 11) fungicides such as Headline and Quadris have lost their efficacy. Ascochyta rabiei, the pathogen causing Ascochyta blight on chickpeas, has developed resistance to these fungicides.
- ProPulse, a new product from Bayer that may be registered as early as 2012, should be used cautiously. ProPulse is a premix of a DMI fungicide (prothioconazole, the active ingredient in Proline) and a carboxamide fungicide (fluopyram). If you apply ProPulse and then Priaxor, Endura, or Vertisan, you will not be rotating fungicide chemistries; likewise, if you apply Proline and then ProPulse, you will not be rotating fungicide chemistries. Carboxamide (FRAC 7) chemistries are high-risk for the development of pathogen resistance, and you need to make sure that you do not make sequential (back-to-back) applications of carboxamide chemistries.

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7. Which fungicides are best?

- Registered fungicides differ in their efficacy against Ascochyta blight on chickpeas.
- Fungicide efficacy testing results have been posted to the NDSU Carrington Research Extension Center website. As you select fungicides, you should assess the performance of fungicides across multiple trials and multiple locations. Fungicide performance can differ in response to which diseases are present, levels of disease when products are applied, environmental conditions, plant architecture and the susceptibility to disease of the chickpea variety planted, crop growth stage at the time of fungicide application, and other factors.

8. Do not plant chickpeas adjacent to a field where chickpeas were recently grown.

Planting chickpeas adjacent to a field where chickpeas were recently grown can result in high Ascochyta disease pressure. The
Ascochyta pathogen produces specialized airborne spores (ascospores) on overwintered diseased chickpea residues, and these
spores are readily carried aloft in air currents. The ascospores are capable of long-distance transport, but adjacent fields receive
particularly high disease pressure.

9. Even if disease-free seed is used and rotations are long, scout your chickpeas carefully for disease.

- The Ascochyta pathogen produces specialized airborne spores (ascospores) on overwintered diseased chickpea residues. These spores are readily carried aloft by air currents and can be moved long distances in the atmosphere. Chickpea fields planted downwind of fields where Ascochyta blight occurred the previous season are at high risk of Ascochyta blight outbreaks caused by these spores.
- In the Pacific Northwest, the release of ascospores from overwintered diseased chickpea residues coincides with emergence and early
 vegetative growth of new chickpea crops: Ascospore release peaks in April and can into May. Ascospores are generally deposited
 onto new crops during rain events.
- Airborne ascospores are an important source of Ascochyta blight in chickpea crops. Even if disease-free seed is used and rotations
 are long, scout your chickpeas carefully for disease.
- Late planting may help chickpeas escape this source of Ascochyta blight, but it is a risky strategy due to the yield penalty associated with late planting and annual fluctuations in the dates of ascospore release.

